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Influence of macerated fenugreek (*Trigonella foenum graecum*) oil added to trout feed at the different rates on the Feed Conversion Rate (FCR), body length, blood parameters and Nitroblue Tetrazolium (NBT) values of rainbow trout (*Oncorhynchus mykiss* Walbaum, 1792)

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**Abstract:** In this study, 1% and 2% of macerated fenugreek oil was added to the feeds of rainbow trout with an average weight of  $25.79 \pm 1.5$  g. At the end of the study, growth rate, blood parameters and NBT (Nitroblue Tetrazolium) level of rainbow trout were determined. The best feed ratio (FCR) was observed in the control group (0.77). Statistically significant differences were found only in MID values (P<0.05), although there was a numerical increase in all blood parameters. There was no statistically significant difference between NBT levels (P> 0.05). Although the best weight gain was in the control group as in the FCR values, the maximum elongation was measured at D1 and then at D2 (P < 0.05). The best survival rate was obtained with 96.66% in D1 while the worst was observed in D2 with 60% (p<0.05).

Key words: Trout; Fenugreek; NBT; Blood parameters; Length parameters.

#### Introduction

Plants are used on fish in various forms as disinfectants or therapeutics in different studies (1, 2, 3, 4, 5). In addition, some researchers have investigated the sensory, chemical, microbial and physiological effects of plants on fish (6, 7). The plants are either given directly to the fish in different ways, or they are left in the water and affect the fish by altering the properties of the water (8). On the other hand, the use of plants in trout breeding as feed additives has been explored for years to ensure healthy and rapid growth of the fish. Most studies do not achieve longitudinal growth, even if the trout are grown to the nearest extent. Providing a balanced growth is the most desirable situation in aquaculture. Fenugreek, also known in traditional Ottoman Medicine, has been used as a height extender (9, 10). Fenugreek contains animal growth stimulants that are not present in other fodder legumes and therefore have the potential to reduce the use of artificial growth promoters. This and other medicinal properties of fenugreek will help reduce the dependence on synthetic drugs that are considered to be serious pollutants of water resources (11).

Fenugreek is a common plant cultured in many parts of Europe, Africa and Asia and it is very rich in terms of steroidal saponins (12). It is also used in traditional medicine due to its lactogenic, anti-diabetic, anti-microbial, anti-cancer and hypocholesterolemic properties (13, 14, 15, 16). Flavonoids and saponins in fenugreek protect the living cells against oxidative stress and stimulate the immune system. It has been reported in some fish studies that fenugreek is activating the immune system of the fish (17, 18, 19).

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Analysis of blood parameters revealed that there is a valuable approach to determine the health status of the cattle raised, and these parameters provide reliable information about metabolic disorders, deficiencies and chronic stress status before being found in the clinical setting (20). Blood biochemistry parameters can be also used to detect the health of fish (21). It has been found that the fenugreek participating in the rasion causes a high amount of hemoglobin, hematocrit, lysozyme and myeloperoxidase activity in fish. It is recommended for feed additives and fish breeding to improve hematological and immune status (22). In addition, fenugreek contains phenolic and flavonoid compounds, thus helping to increase antioxidant capacity in the liver (23).

In this study, we aimed to determine the feed rate (FCR), some blood and length parameters and NBT values of the rainbow trout (*O. mykiss*) of macerated fenugreek oil added to rasion at different ratios.

#### **Materials and Methods**

# Material

#### **Experimental materials**

Rainbow trout (*O. mykiss*) with an average live weight of  $25.79 \pm 1.5$  g was used in the study. Fenugreek (*T. foenum graecum*) was kept in 1/10 sunflower oil for 15 days to obtain macerated oil. The obtained macerated oil was added to a commercial trout diet at

Fahle 1	Mean	values	of blood	narameters	measured	at the	end of 21	days fe	eding	according	to exi	nerimental	orom	ns
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	WBC	LYM	MID	GRAN	RBC	HGB	HCT	MCV	MCH	MCHC	RDW-SD
	(10 <sup>3</sup> /µL)	(10 <sup>3</sup> /µL)	(10 <sup>3</sup> /µL)	(10 <sup>3</sup> /µL)	(10 <sup>6</sup> /µL)	(g/dl)	(%)	fL	pg	g/dl	(fL)
С	47,57	44,4	1,84	0,97	1,25	6,87	16,24	129,85	55,55	42,88	64,78
D 1	54,48	50,6	2,58	1,28	1,65	9,11	21,94	133	54,98	41,42	66,92
D 2	52,77	49,15	2,42	1,17	1,51	8,21	19,71	132,5	54,11	41,54	69,08

1% and 2% by spraying method. The study groups were formed as control (C) (no fenugreek oil added to the diet), experiment 1 (D1) (1% macerated fenugreek oil added to the diet) and experiment 2 (D2) (2% macerated fenugreek oil added to the diet). Ten fish were studied in each group and three were repeated. Fishes are fed daily with feed at a rate of 2% of their body weight. Feeding was done daily for 21 days, morning and evening. Feed evaluation ratio (FCR) and mean height values were determined at the end of the study.

Fishes were anesthetized (Benzocaine 30 mg/L) before the blood collection. Experiments were conducted in accordance with ethical rules (Inonu University, Faculty of Medicine, Experimental Animal Ethics Committee, Protocol No:2017/A-24). Blood samples were taken from the tail veins of the fried fishes by injector and transferred to tubes containing EDTA. On the same day nitro-blue tetrazolium (NBT) activity (total oxidative radical production of neutrophils) from EDTA blood samples was determined spectrophotometrically. Full blood count was also performed with PROCAN PE-6800VET brand fully automated hematology analyze.

The measured water in the experiment was determined as temperature 14.61 °C, pH 8.25, Conductivity 16.9 (dS/m), Hardness 11.25 A ° S, Salinity 8.4 mg/l.

# Methods

#### Analysis of hematologic parameters

In the 21<sup>st</sup> days of the application, the blood samples of fish (n=10) randomly selected from all groups were taken. Fish were anesthetized with the 0.30 ml L<sup>-1</sup> 2-phenoxyethanol for 10 minutes (24) and observed anesthesia of fish under deep sedation, including loss of swimming action and partial loss of equilibrium (25). The blood samples were taken from the caudal vena into K2 EDTA tubes (4 ml). The HCT, Hg, RBC, WBC, mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), mean corpuscular volume (MCV), and the activity of Nitroblue-Tetrazolium (NBT) in the blood samples and analysed by fully automatic hematology analyser (PROCAN PE-6800VET Brand) in the same day.

# Analysis of Nitroblue Tetrazolium (NBT) activity

The 0.1 ml of 0.2 % NBT solution was added to the microtiter plates and an equal amount of blood was added on it. After this suspension was allowed to stand at room temperature for 30 minutes, 0.05 ml of it was added to glass tubes containing 1 ml of N, Ndimethylaformamide. After centrifugation at 3000 rpm for 5 minutes (centrifuge; Nuve Brand NF800 Model), the upper layer was read on a spectrophotometer (Hach Lange Brand DR6000 Model) at 540 nm.

# Length, weight measurements and determination of FCR values

The length of the fish was measured with a ruler and their weights were weighed on the precision scale. Feed Conversion Ratio was obtained by the following formula:

F.C.R. = Feed given / Animal weight gain

### Statistical analysis

The data obtained were statistically tested using the SPSS statistical program at the 0.05 confidence interval by DUNCAN test (P<0.05). The mean values were given in the results as mean  $\pm$  standart error of means. However, all results were analysed by the SPSS 24.0 Package Programme.

### Results

### Hematological parameters

A numerical increase in all blood parameters outside the MCHC was observed for fish in the D1 and D2 groups. On the other hand, only a statistically significant difference was found in the MID (Monocyte) parameters (P<0,05). No statistically significant difference was found in other blood parameters (P>0,05). The blood parameters measured at the end of the 21-day trial are given in Table 1.

# Nitroblue Tetrazolium (NBT) activity

NBT values were measured as  $0.296 \pm 0.09$  before study in the control group. At the end of the 21 day study;  $0.353 \pm 0.06$  in the control group,  $0.332 \pm 0.03$  in the D1 and  $0.305 \pm 0.05$  in the D2. Although the obtained data show an increase compared to the values obtained before feeding, NBT activity was not significant in the case of macerated oil application due to its water-soluble phenolic compounds in fenugreek.

#### Feed conversion ratio

In this study, the best FCR value was obtained in the control group. Although there were numerical differences between the control and the two experimental groups, no statistically significant difference was observed (P>0.05). Table 2 shows FCR change values.

#### Length increase

The best elongation value at the end of 21 days of feeding was obtained at D1. A statistically significant difference was found between the length of the fish measured before the feeding and the length of the fish fed with the fenugreek supplemented feed (P>0.05)

**Table 2.** FCR, survival rates, mean weight and mean length change values.

	Control	D 1	D 2
FCR	0,77	1,14	1,04

Table 3. Mean	length and	mean weight	change values.
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-	-	-	
	Control	D 1	D 2
Lenght 0 day (Average±sd)	13.08±1.5	13.02±1.4	12.94±1.9
Lenght 21 day (Average±sd)	13.68±1.6	14.39±1.4	14.30±1.9
Average Weight 0 Day	27.29±1.0	24.73±1.2	25.35±1.1
Average Weight 21 Day	40.49±1.3	35.76±1.1	34.72±1.3

Table 4. Survival rate.

	Control	D 1	D 2
Survival Rate %	73.33	96.66	60

(Table 3).

#### Survive rate

The best survival rate was obtained with 96.66% in D1 while the worst was observed in D2 with 60% (P<0.05) (Table 4).

### Discussion

In our study, the increase in MCV values of the fenugreek applied groups was higher in the 1% group, but both groups showed an upward image. The MCV increase is a macrocytosis indicator with a decrease in the MCHC value in both treatment groups. This is the evidence that fenugreek oil stimulates renewal and the production of new erythrocytes. The parallel increase in MCV values of lenght values revealed that oxygenation and blood production were increased by stimulation. In another study of fenugreek applied on Sparus aurata L., the increases in RBC value were reported to increase in all fenugreek applied groups, although not statistically significant, as in our study (26). Yilmaz et al. (27) found that there was no significant change in the RBC, HGB, HCT values of *Dicentrarchus labrax*. In the same way Roohi et al. (28) did not observe a statistical change in these values of carp.

In addition to this, another study done by Gültepe et al. (29) found an increase in RBC, HGB, HCT values of fenugreek applied tilapia. A similar study by Acar et al. (30), found an increase in RBC and HGB values.

Monocytes are groups of cells that increase at the time of chronic stress (31, 32). In our study, the monocyte levels in the fenugreek oil application groups were higher than the control group. Monocyte levels in phagocytic and lysosomal function-induced immune systems are as high as in our study groups (29, 33). Monocyte increase, one of the representatives of the cellular immune response, was also found to be high in other studies performed with fenugreek (34). In another study where fenugreek oil was applied, it was found that the number of monocytes increased. This state that the substances contained in the fenugreek oil suppress ACE (The enzyme that enables the conversion of angiotensin 1 to angiotensin 2 is a vasoconstrictor) activity (35). This suppression increases the nutrient and oxygenation levels of the tissues by increasing vessel expansion.

The RDW-SD value was found to be higher in the two experimental groups than in the control. This elevation is indicative of the number of large erythrocytes, indicating that the fenugreek oil increases the formation



of new erythrocytes. Take Güllü et al. (36) in a similar experiment found that RBC, HGB and HCT values of the group fed with fenugreek-supplemented diet were not different from those of the control group.

In this study, feed conversion rate in experimental groups decreased compared to control group. There was no statistically significant difference between FCR values (Control: 0.77; D1: 1.14; D2: 1.04) in this study. (P>0.05). Lawal et al. (37), In an FCR study on fenugreek-fed tilapia fish, no significant difference was found between the experimental groups and the control group. In a study on Cyprinus carpio, it was observed that the rate of feed evaluation decreased in a similar way to this study (28). Öz et al. (38) investigated the effect of black black cumin oil on trout. It has been observed that the feed conversion rate of trout fed with feed of black cumin oil addition increased feed consumption and daily feed consumption of fish for each.

Araee et al. (39) reported that fenugreek rats were effective on bone marrow haematopoietic stem cells, In this study, fish fed with fenugreek oil extended their length more than control group. In the experimental groups, the weight values were less increased compared to the control group, on the contrary, the lengths of the experimental groups were increased more than the control groups.

NBT activity was not significant in the case of macerated oil application due to its water-soluble phenolic compounds in fenugreek. Kumar et al. (40) found that NBT activity was at the highest level in a study used by fenugreek.

As a result, it is possible to say that the addition of fenugreek macerated oil to trout feeds gives positive results. Macerated fenugreek oil reveals numerical differences in all parameters. However, the most statistically significant result was that survival rates at Experiment Group 1 and fish fed with fenugreek fat-fed diet had a longer prolongation of their size. It may be possible to obtain more striking results by developing this research in future studies.

## Authors' contributions

All of the responsibilities and contributions belong to correspounding author.

# **Conflict of interest disclosure**

There is no conflict of interest of this manuscript.

# References

1. Altınterim B, Öztürk E, Kutluyer F, Aksu O. Yeşil Çay Yağının Gökkuşağı Alabalıklarının (Oncorhynchus mykiss) Yem Değerlendirme Oranına ve Hematolojik Parametrelerine Etkileri. Atatürk Üniversitesi Veteriner Bilimleri Dergisi, 2018a; 13(2): 159-164..

2. Chorianopoulos NG, Giaouris ED, Skandamis PN, Haroutounian SA. and Nychas, GJE. Disinfectant test against monoculture and mixed-culture biofilms composed of technological, spoilage and pathogenic bacteria: Bactericidal effect of essential oil and hydrosol of *Satureja thymbra* and comparison with standard acidbase sanitizers. Journal of Applied Microbiology, 2008; 104: 1586–1596.

3. Chang ST, Chen PF and Chang SC. Antibacteriel activity of leaf essantial oils and their consituents from *Cinnamon osmophloeum*. Journal of Ethanophermacology, 2001; 77: 123-127.

4. Gulec AK, Altinterim B. and Aksu O. Determination of lethal concentration (LC50) values of *Cinnamomum zeylanicum* hydrosol on carp fish. Iranian Journal of Fisheries Sciences, 2013; 12(1): 34-44

5. Altinterim B, Kutluyer F. and Aksu, O. Effects of different plant oils having different oxygen radical absorbance capacity (ORAC) on hematological parameters of rainbow trout (*Oncorhynchus mykiss*) at high stocking density. Ataturk University Journal of Veterinary Sciences, 2018b; 13(1): 63-69.

6. Öz M. Effects of garlic (Allium sativum) supplemented fish diet on sensory, chemical and microbiological properties of rainbow trout during storage at -18 °C. LWT, 2018; 92: 155-160.

7. ÖZ M. Effects of black cumin (Nigella sativa) oil on ammonia and biogenic amine production in rainbow trout. Indian J. Anim. Res, 2018; 52(2): 265-269.

8. Altınterim B, Gulec AK and Aksu O. Determination of Safety Dose of *Eucalyptus camaldulensis* Hydrosol on Mirror Carp (*Cyprinus carpio*). Fresenius Environmental Bulletin, 2012; 21(5a): 1219-1222.

9. Altınterim B, Danabas D. and Aksu O. The effects of common yarrow (Achillea millefolium Linnaeus), cinnamon (Cinnamomum zeylanicum Blume) and rosemary (Rosemarinus officinalis Linnaeus) hydrosols on the some immunological and hematological parameters of common carp (Cyprinus carpio L., 1758). Cellular and Molecular Biology, 2018c; 64(14):19-24.

10. Telli B. Kenzü's-Sıhhatü'l-Ebdâniyye Eser-İ Mürşîd-İ Osmâniyye'de Bitki Adları. Kesit Akademi Dergisi (The Journal of Kesit Academy), 2015; 1: 96-116.

 Acharya SN, Thomas JE and Basu SK. Fenugreek: an "old world" crop for the "new world". Biodiversity, 2006; 7(3-4): 27-30.
 Petropoulos GA. Fenugreek–Thegenus Trigonella. 1st Edn., Taylor and Francis, Londonand New York, 2002; Pp: 200.

13. Wani SA, Kumar P. Fenugreek: A review on its nutraceutical properties and utilization in various food products. J. Saudi Soc. Agric. Sci., 2018; 17:97–106.

14. Kim JK, Park SU. An update on the biological and pharmacological activities of diosgenin. Excli J., 2018; 17:24–28.

15. Bahmani M, Shirzad H, Mirhosseini M, Mesripour A, Rafieian-Kopaei M. A review on ethnobotanical and therapeutic uses of Fenugreek (*Trigonella foenum-graceum* L.) J. Evid. Based Complement. Altern. Med., 2016; 21:53–62.

16. Srinivasan K. Fenugreek (*Trigonella foenum-graecum*): A review of health beneficial physiological effects. Food Rev. Int., 2006; 22:203–224.

17. Rababah TM, Hettiarachchyand NS and Horax R. Total phenolics and antioxidant activities of fenugreek, gren tea, black tea, grape seed, ginger, rosemary, gotu kola, and ginkgo extracts, vitamin E, and tert butylhydroquinone. Journal of Agriculture, Food and Chemistry, 2004; 52: 5183 - 5186.

 Shalaby SMM. Response of Niletilapia (*Oreochromis niloticus*) fingerlings to diets supplemented with different levels of fenugreek seeds (Hulba). Journal of Agriculture Mansoura University, 2004; 29: 2231-2242

19. Mustafa AAZM, Ahmad MH, Mousallamy A and Samir A. Effect of Using Dried Fenugreek Seeds as Natural Feed Additives on Growth Performance, Feed Utilization, Whole-body Composition and Entro-pathogenic *Aeromonas hydrophila* challenge of Monosex Nile Tilapia *O. niloticus* (L) Fingerlings. Australian Journal of Basic and Applied Sciences, 2009; 3(2): 1234-1245.

20. Bahmani M, Kazemi R and Donskaya P. A comparative study of some hematological features in young reared sturgeons (*Acipenser persicus* and *Huso huso*). Fish Physiol Biochem, 2001; 24: 135–140. 21. De Pedro N, Guijarro AE, Lopez-Patino MA, Marinez-Alvarez R and Delgado M. Daily and seasonal variation in haematological and blood biochemical parameters in tench *Tinca tinca*. Aquaculture Res, 2005; 36: 85–96.

22. Yilmaz S, Ergun S and Yigit M. 2011. Effects of thyme, rosemary and fenugreek on some hematological and immunological parameters of tilapia, *Oreochromis mossambicus*, Aquaculture. Europe, Conferince 18- 21 October 2011, Rhodes, Greece.

23. Dixit P, Ghaskadbi S, Mohan H and Devasagayam TP. Antioxidant properties of germinated fenugreek seeds, Phytotherapy Research, 2005; 19(11): 977-983.

24. Velisek J, Svobodova Z. Anaesthesia of common carp (*Cyprinus carpio* L.) with 2-phenoxyethanol: Acute toxicity and effects on biochemical blood profile. Acta Vet. Brno, 2004; 73:247-252.

25. Altun T, Danabas D. Effects of short and long exposure to the anesthetic 2-phenoxyethanol mixed with ethyl alcohol on common carp (*Cyprinus carpio* L., 1758) fingerlings. Isr. J. Aquacult.—Bamidgeh, 2006; 58(3):1–5.

26. Awad E, Cerezuela RM and Esteban A. 2015. Effects of fenugreek (*Trigonella foenum graecum*) on gilthead seabream (*Sparus aurata* L.) immune status and growth performance. *Fish and Shell-fish Immunology*, 45: 454-464.

27. Yilmaz S, Ergun S and Celik ES. Effect of dietary spice supplementations on welfare status of sea bass, *Dicentrarchus labrax* L. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences, 2016; 86: 229–237.

28. Roohi Z,Imanpoor MR, Jafari V and Taghizadeh V. The use of fenugreek seed meal in fish diets: growth performance, haematological and biochemical parameters, survival and stres resistance of common carp (*Cyprinus carpio* L.). Aquaculture Research, 2015; 48: 1209–1215.

29. Gültepe N, Bilen S, Yılmaz S, Güro, D and Aydın S. Effects of herbs and spice on health status of tilapia (*Oreochromis mossambicus*) challenged with *Streptococcus iniae*. Acta Veterineria Brno, 2014; 83: 125–131.

30. Acar Ü, Kesbic OS, Yilmaz S, Gultepe N and Turker A. Evaluation of the effects of essential oil extracted from sweet orange peel (*Citrus sinensis*) on growth rate of tilapia (*Oreochromis mossambicus*) and possible disease resistance against *Streptococcus iniae*. Aquaculture, 2015; 437: 282-286.

31. Baldisserotto B, Romero JMM and Kapoor B. Fish osmoregulation. CRC Press/ Taylor and Francis Group, Boca Raton, 2007; Pp 540.

32. Mazon A, Monteiro E, Pinheiro G and Fernadez M. Hematological and physiological changes induced by short-term exposure to copper in the freshwater fish, *Prochilodus scrofa*. Brazilian Journal of Biology, 2002; 62: 621–631.

33. Joo HS, Kalbassi MR and Johari SA. Hematological and histo-

pathological effects of silver nanoparticles in rainbow trout (*Onco-rhynchus mykiss*)-how about increase of salinity? Environmental Science and Pollution Research, 2018; 25: 15449–15461.

34. Emeish WFA. and El-Deen AGS. Immunomodulatory Effects Of Thyme and Fenugreek in Sharp Tooth Catfish, *Clarias gariepinus*. Assiut Veterinary Medical Journal, 2016; 62(150): 1-7.

35. Hamden K, Keskes H, Belhaj S, Mnafgui K, Feki A and Allouche N. Inhibitory potential of omega-3 fatty and fenugreek essential oil on key enzymes of carbohydrate-digestion and hypertension in diabetes rats. Hamden et al. Lipids in Health and Disease, 2011; 10: 226.

36. Güllü K, Acar Ü, Kesbiç OS, Yılmaz S, Ağdamar S, Ergün S and Türker A. Beneficial effects of Oral Allspice, *Pimenta dioica* powder supplementation on the hematoimmunological and serum biochemical responses of *Oreochromis mossambicus*. Aquaculture Research, 2016; 47: 2697–2704. 37. Lawal MO, Aderolu AZ and Aarode OO. 2013. Growth and economic performance of Nile Tilapia, *Oreochromis niloticus* (L.) fingerlings fed diets containing graded levels of sclerotium. AACL Bioflux, 2013; 6(3): 180-187.

38. ÖZ M., Dikel, S, and Durmus M. Effect of black cumin oil (Nigella sativa) on the growth performance, body composition and fatty acid profile of rainbow trout (Oncorhynchus mykiss). Iranian Journal of Fisheries Sciences, 2018; 17(4): 713-724.

39. Araee M, Norouzi M, Habibi G And Sheikhvatan M. Toxicity of *Trigonella foenum graceum* (Fenugreek) in bone marrow cell proliferation in rat. Pakistan Journal of Pharmaceutical Sciences, 2009; 22(2):126-130.

40. Kumar A, Vasmatkar P, Baral P, Agarawal S And Mishra A. Immunomodulatory and growth promoting effect of dietary fenugreek seeds in fingerlings of common carp (*Cyprinuscarpio* Lin.). Fishery Technology, 2017; 54: 170-175.